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Journal of Coordination Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713455674

¹⁵N AND ³¹P NMR STUDIES OF CYANO(TRIALKYL/TRIARYL)PHOSPHINE GOLD(I) COMPLEXES

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To cite this Article Akhtar, M. Naseem, Gazi, Ibrahim H., Isab, Anvarhusein A., Al-Arfaj, Abdul Rahman, Wazeer, Mohammed I. M. and Hussain, M. Sakhawat(1995) '¹⁵N AND ³¹P NMR STUDIES OF CYANO(TRIALKYL/TRIARYL)PHOSPHINE GOLD(I) COMPLEXES', Journal of Coordination Chemistry, 36: 2, 149 – 157

To link to this Article: DOI: 10.1080/00958979508022555 URL: http://dx.doi.org/10.1080/00958979508022555

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¹⁵N AND ³¹P NMR STUDIES OF CYANO(TRIALKYL/TRIARYL)PHOSPHINE GOLD(I) COMPLEXES

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(Received December 19, 1994; in final form May 2, 1995)

The ligand scrambling reaction of $R_3PAuC^{15}N$ to form $(R_3P)_2Au^+$ and $Au(C^{15}N)_2^-$ has been studied (by ¹⁵N and ³¹P NMR spectroscopy) for R = Me, Et, *i*-Pr, and Ph. ³¹P NMR showed two resonances due to R_3PAuCN and $(R_3P)_2Au^+$ species, while ¹⁵N NMR showed only an averaged resonance due to $R_3PAuC^{15}N$ and $Au(C^{15}N)_2^-$ species, except for Et₃PAuC¹⁵N, for which two separate resonances were detected. $R_3PAu^{13}C^{15}N$ (where R = Me, Et and Ph) complexes were also prepared and ²J(³¹P-¹³C) as well as ³J(³¹P-¹⁵N) constants were measured. The free activation energy for ligand scrambling in Ph₃PAuCN was determined by ³¹P NMR band shape analysis to be 39.7 kJ/mol⁻¹.

KEYWORDS: gold, phosphines, cyanide, nmr, scrambling

INTRODUCTION

Recent solid state studies¹⁻³ of cyano(trialkyl)phosphinegold(I) complexes have shown that they form linear gold(I) species. However, in solution, these complexes undergo a ligand scrambling reaction as shown in (I).^{2,4}

$$2R_3PAuCN \rightleftharpoons (R_3P)_2Au^+ + Au(CN)_2^-$$
(1)

The equilbrium constant for these reaction is dependent⁴ upon extrinsic effects such as initial concentration of the complexes, $[R_3PAuCN]_0$, and ionic strength of the medium, and intrinsic factors such as the steric effects and electronic properties of the phosphine ligands. These ligand scrambling reactions are important from a biological point of view. Smokers who are treated with various anti-arthritic gold drugs were reported to have higher concentration of gold in their red blood cells as compared with nonsmokers.⁵⁻⁷ Possible mechanisms by which the gold in the form of Au(CN)²/₂ can enter into red blood cells is the subject of several recent studies.⁸⁻¹² The very large formation constant for Au(CN)²/₂ (reported to be log β_2 36.6,¹³) drives all L-Au-CN complexes (where L = thiolate- or phosphine-containing

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ligands) to disproportionate as in (1) thus generating $Au(CN)_2^-$ which can enter into red blood cells. Thiolate-based drugs do not enter red blood cells because they are in a polymeric form.^{11,14,15}

We have recently reported the disproportionation of gold(I)-thiomalate and gold(I)-thioglucose in the presence of $C^{15}N^-$ by using ¹⁵N NMR spectroscopy.¹⁶ At 1:<2 AuSR:C¹⁵N⁻ ratio, two resonances were observed for Au(C¹⁵N)² and RS-Au-C¹⁵N⁻. At a 1:2 ratio of AuSR:C¹⁵N⁻, only one resonance was observed for Au(C¹⁵N)² complexes, which means that all the thiolate ligands are released in solution. In the present study, we have synthesised ¹⁵N labelled R₃PAuC¹⁵N complexes to see if the ligand scrambling reaction can be followed by ¹⁵N NMR spectroscopy and also to compare ¹⁵N chemical shifts of RS-Au-C¹⁵N⁻ with R₃P-Au-C¹⁵N complexes. We have also prepared doubly labelled R₃P-Au-¹³C¹⁵N (where R = Me, Et and Ph) complexes, where all relevant coupling constants were measured.

EXPERIMENTAL

Reagents

 $KC^{15}N$ (99%) and $K^{13}C^{15}N$ (99%) were purchased from Merck Sharp and Dohme, Canada. Sodium tetrachloroaurate dihydrate, phosphine ligands (trimethyl, triethyl, triphenyl, triisopropyl) were obtained from Strem Chemical Co. CD_3OD was obtained from Fluka Chemical Co.

R ₃*PAuCl*

All the R_3PAuCl complexes were prepared by the addition of phosphine ligands to an ethanolic solution of NaAuCl₄ as described in the literature.^{17–18}

$R_{3}PAuC^{15}N$

Tri(alkyl/aryl) phosphinegold(I) cyanide complexes were prepared by adding solid $KC^{15}N$ (10% labelled) directly to an alcoholic solution of R_3PAuCl . Elemental analysis, melting points and percentage yields for $R_3PAuC^{15}N$ (R = Me, Et, *i*-Pr, Ph) complexes are given in Table 1.

R	Found (calcd.)%			Found(lit.)	% yield
	C	Н	N	m.pt.	
Me	15.94	2.97	3.86	201	80
	(16.0)	(3.0)	(5.0)		
Et	25.08	4.54	4.16	109 (113) ^a	41
	(24.58)	(4.39)	(4.39)	· · ·	
i-Pr	31.64	5.77	3.80	108	54
	(31.27)	(5.47)	(3.91)		
Ph	46.23	3.29	2.81	209 (204) ^a	90
	(46.93)	(3.08)	(3.08)		

Table 1 Elemental analyses of R₃PAuC¹⁵N complex, melting points and % yield.

^aReferences 2 and 4.

$R_3 PAu^{13}C^{15}N$

These complexes were synthesized as decribed above (R = Me, Et and Ph) using 99% label ¹³C and 99% ¹⁵N label K¹³C¹⁵N.

¹⁵N and ³¹P NMR Spectroscopy

¹⁵N NMR spectra were obtained at 27.24 MHz on a Jeol 270 spectrometer using CD₃OD solutions at the probe temperature of 300 K. The chemical shifts for ¹⁵N NMR spectra were measured using a sealed $\rm NH_4^{15}NO_3$ solution as external reference, which has a resonance at 375.11 ppm relative to pure CH₃NO₂ 380.2 ppm.^{16,19}

We did not measure the T_1 values of any resonance. The spectrometer conditions were: 5.0 s delay time, 16 K data points, aquisition width 20,000 Hz, pulse width 6.0 μ s (20°), and 10 mm multinuclear probe. For the ¹⁵N NMR spectra, approximately 60,000 scans were accumulated for each sample.

³¹P NMR spectra were obtained at 109.25 MHz using the above solutions at the probe temperature of 300 K. The ³¹P NMR chemical shifts were measured against external 1% TMP.²⁰ T₁ values were previously reported^{2,4} for these compounds and are in the range of 5–7 seconds. The spectrometer conditions were: 1.0 s delay time, 32 K data points, acquisition width 4,000 Hz, pulse width 5.0 μ s, 10 mm multinuclear probe. For the ³¹P NMR spectra, approximately 10,000 scans were accumulated. ¹⁵N and ³¹P NMR chemical shifts for various R₃PAuC¹⁵N complexes are given in Table 2, and ²J and ³J values are given in Table 3.

Calculations of Activation Parameters

The activation parameters were calculated from proton decoupled ³¹P NMR spectra for Ph₃PAuCN (0.010 M in CD₃OD), recorded between +25°C and -60°C. At -60°C, two peaks due to Ph₃PAuCN and (Ph₃P)₂Au⁺ are observed and these coalesce around -25°C. Above -20°C, the average peak becomes sharper. At temperatures below - 30°C, integration of ³¹P resonances gave relative concentrations of Ph₃PAuCN and (Ph₃P)₂Au⁺ species and from this K_{eq} was calculated. K_{eq} was found to be constant at 0.14 for -60°C, -50°C and -40°C, within experimental error.

Band shape analysis was carried out on the basis of exchange between two non-coupled sites with unequal population, using a library package.²¹ The life time and rate constant at different temperatures are given in Table 4. Activation energies

R	$\delta(^{15}N)^{b}$ R ₃ PAuCN	$\delta(^{13}C)^d$ R ₃ PAuCN	$\delta(^{31}P)^{c}$ R ₃ PAuCN	$\delta(^{31}P)$ (R3P) ₂ Au ⁺	(K _{eq}) ^d	v _{co} ,cm ^{-1e}
Me	263.0	158.3	- 3.09	5.94	0.37	2064
Et	262.5	160.4	35.35	44.15	0.24	2062
<i>i</i> -Pr	262.2	160.9	66.10	75.20	0.29	2059
Ph	265.5	156.2	39.37	43.92	0.14	2069

Table 2 ¹³C, ¹⁵N and ³¹P NMR chemical shifts and K_{eo} data for R₃PAuC¹⁵N complexes.^a

 $^{a}CD_{3}OD$ solvent. ^{b}In ppm relative to external NH₄ $^{15}NO_{3}$. ^{c}In ppm relative to external 1% TMP. $^{d}Reference 4$. $^{e}Reference 24$.

Compound	Temp. (K)	² J (³¹ P- ¹³ C) (Hz)	³ J (³¹ P- ¹⁵ N) (Hz)
Me ₃ PAu ¹³ C ¹⁵ N	297	130.7(129.4) ^a	3.6
$Et_3 PAu^{13}C^{15}N$	297	122.2(122.2) ^a	2.9
Ph ₃ PAu ¹³ C ¹⁵ N	213	124.6(126.0) ^a	4.0

Table 3 ³¹P Coupling constant data for $R_3PAu^{13}C^{15}N$, where R = Me, Et, and Ph.

^aReference 4.

Table 4 Life time^a and rate constant data for the scrambling reaction of 0.010M Ph₃PAuCN in CD₃OD at $K_{eq} = 0.14$.

$(\tau_a/s)^a$	k/dm ³ mol ⁻¹ s ⁻¹	
0.0079	1840	
0.0024	6056	
0.00063	23.071	
0.00030	48,450	
0.00012	121,124	
	$\begin{array}{c} (\tau_a/s)^a \\ \hline 0.0079 \\ 0.0024 \\ 0.00063 \\ 0.00030 \\ 0.00012 \end{array}$	

^aLife time of the Ph₃PAuCN species.

were calculated from an Arrthenius plot of $\ln k_r$ versus 1/T and free energy of activation from an Eyring plot of $\ln (k_r/T)$ versus 1/T.

RESULTS AND DISCUSSION

³¹P NMR spectra of 0.020 M Me₃PAuCN, Et₃PAuCN, *i*-Pr₃PAuCN and Ph₃PAuCN complexes in CD₃OD were measured. In each case two resonances assigned to R₃PAuCN and (R₃P)₂Au⁺ were observed as indicated in (*1*) except for Ph₃PAuCN. Note that the ratios of R₃PAuCN and (R₃P)₂Au⁺ resonances are different because K_{eq} is different for each complex,⁴ as shown in Table 2.

Figure 1A, 1B and 1C shows the ¹⁵N NMR spectra of Me₃PAuC¹⁵N, Et₃PAuC¹⁵N, and *i*-Pr₃PAuC¹⁵N complexes using the same sample as for the ³¹P NMR measurements. The ¹⁵N chemical shift difference between Me₃PAuC¹⁵N or *i*-Pr₃PAuC¹⁵N and Au(C¹⁵N)² is very small and therefore only one set of resonances is observed. However, two resonances are observed for the Et₃PAuC¹⁵N complex. The less intense resonance at 264.0 ppm is due to Au(C¹⁵N)² and the more intense resonance at 262.5 ppm is assigned to Et₃PAuC¹⁵N.

Figure 2 shows ³¹P NMR spectra of $R_3PAu^{13}C^{15}N$ (R = Me, Et and Ph) complexes. Both Me₃PAu¹³C¹⁵N and Et₃PAu¹³C¹⁵N gave two sets of peaks due to $(R_3P)_2Au^+$ and $R_3PAu^{13}C^{15}N$ which split into two resonances due to ²J(³¹P-¹³C) coupling.^{2,4} The ²J values are given in Table 3. However, Ph₃PAu¹³C¹⁵N gave only one average resonance due to fast exchange between $(Ph_3P)_2Au^+$ and $Ph_3PAu^{13}C^{15}N$ species at room temperature, consistent with previous studies.⁴ On cooling to $-60^{\circ}C$, all species were in the slow exchange limit and resonances due to both Ph₃PAu¹³C¹⁵N and $(Ph_3P)_2Au^+$ were observed as shown in Figure 2. ³J splitting for $R_3PAu^{13}C^{15}N$ due to ³¹P-¹⁵N was also observed as shown in Figure 2 and reported in Table 3.

The X-ray structure of Me₃PAuCN, Et₃PAuCN, and Ph₃PAuCN shows that these complexes are linear in the solid state.^{1–3} However, as soon as they are dissolved in solution,^{2,4,22} these complexes undergo ligand scrambling reactions as shown in (1).



Figure 1 The 27.24 MHz ¹H noise-decoupled ¹⁵N NMR spectra in CD₃OD of [A] 0.020 M $Me_3PAuC^{15}N$, [B] 0.020 M $Et_3PAuC^{15}N$, [C] 0.020 M *i*-Pr₃PAuC¹⁵N.

The previously recorded⁴ K_{eq} values of these complexes are given in Table 2. K_{eq} is different for all complexes under investigation and this suggests that various factors influence K_{eq} . These have been studied by Horman-Arendt and Shaw and include solvent, size of the ligands, initial concentration of complexes, ionic strength of the medium, *etc.*^{2,4}

Unfortunately, the ¹⁵N chemical shift differences between $Au(C^{15}N)_2^-$ and $R_3PAuC^{15}N$ are very small and not resolved, except for $Et_3PAuC^{15}N$. The





Figure 2 The 109.25 MHz ¹H noise-decoupled ³¹P NMR spectra in CD₃OD of: [A], (I) 0.020 M Me₃PAuCN, (II) 0.020 M Me₃PAu¹³C¹⁵N, (III) expanded spectrum of II (all spectra are recorded at 297 K); (B) (I) 0.020 M Et₃PAuCN, (II) 0.020 M Et₃PAu¹³C¹⁵N, (III) expanded spectrum of II (all spectra are recorded at 297 K); (B) (I) 0.020 M Et₃PAu¹³C¹⁵N, (III) expanded spectrum of II (all spectra are recorded at 297 K); (C] (I) 0.010 M Ph₃PAu¹³C¹⁵N at 233 K, (III) expanded spectrum of II.

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assignment of Au($C^{15}N$)₂ and Et₃PAu $C^{15}N$ resonances is based on the K_{eq} value of the complex,⁴ where the Au($C^{15}N$)₂⁻ resonance has to be less intense than that of Et₃PAuC¹⁵N. The broad ¹⁵N resonance for Me₃PAuC¹⁵N (Fig. 1A) indicates that Au $(C^{15}N)_2^{-1}$ and Me₃PAu $C^{15}N$ are in slow exchange. However, for *i*-Pr₃PAu $C^{15}N$, the ¹⁵N resonance is sharp, indicating fast exchange. For Ph₃PAuC¹⁵N (not shown), the exchange-averaged ¹⁵N resonance was observed at 265.5 ppm.

Recently, we studied the interaction of $C^{15}N^-$ with gold(I)-thiomalate (Autm) and gold(I)-thioglucose (Autg) in D₂O at pH* 7.40, using ¹⁵N NMR which showed the presence of Au(C¹⁵N)₂ and RSAuC¹⁵N⁻ (where RS = tm⁻ or tg⁻) at 265.94 ppm and 260.30 ppm, respectively.¹⁶ It should be noted here that for both tm-AuC¹⁵N⁻ and tg-Au-C¹⁵N⁻ species, the resonances appeared at 260.30 ppm. In the present study, the ¹⁵N chemical shift range for all the R₃PAuC¹⁵N species is small, between 262.2 to 265.5 ppm in CD₃OD. As noted in Table 3, ²J and ³J values for Me₃PAu¹³C¹⁵N are greater than for



Figure 3 Dependence of the (A) 13 C and (B) 15 N chemical shifts of R₃PAu¹³CN (or R₃PAuC¹⁵N) complexes vs v(CO). The solid lines are regression lines. The 13 C chemical shift value are taken from ref. 4 and v(CO) values from ref. 24.

 $Et_3PAu^{13}C^{15}N$ at 297 K; a similar observation was made for $R_3PAu(^{15}N-phthalimide)$ (where R = Me, Et, *i*-Pr, Ph, *etc.*) complexes.²³ The magnitude of the ²J and ³J coupling constants is expected to reflect the strength of the P-Au bond. For example, increasing the cone angle decreases strength of the P-M bond.

The electronic parameter, v(CO) for $R_3PNi(CO)_3$, is used by Tollman²⁴ to demonstrate the relative basicity of phosphines. A decrease in v(CO) indicates a net increase in the electron-donating ability of a phosphine. Figure 3 shows a linear correlation between v(CO) vs¹⁵N chemical shifts of the $R_3PAuC^{15}N$ complexes. As shown in Table 2 and discussed by Hormann-Arendt and Shaw,⁴ it is clear that K_{eq} for the Ph₃PAuCN is less than for alkyl R_3PAuCN complexes. There is an inverse correlation between K_{eq} and v(CO).⁴

correlation between K_{eq} and v(CO).⁴ Brown *et al.*^{25,26} studied the IR, and ¹³C and ¹⁵N NMR chemical shifts for ¹⁵N¹³CCo(D₂H₂)L where L = pyridine, primary amine and 4-substituted aniline and their analogues. Interestingly, ¹⁵N¹³CCo(D₂H₂)L and R₃PAu¹³C¹⁵N complexes show an inverse relationship between ¹³C vs ¹⁵N NMR chemical shifts (Figures 3 and 4). This could be due to the metal-to-carbon d_π-p_π electron donation, which will lower the charge on the metal; consequently the electron density will be transferred from a filled metal *d*-orbital into the carbon-centred member of the cyanide π^* -antibonding orbital. Most of the electron density donated from the metal to the cyanide ligand would be expected to reside on the cyanide carbon, and this would cause a ¹³C chemical shift down field and an upfield shift of the ¹⁵N resonance.

 $\Delta G^{\#}$ for the scrambling reaction of Ph₃PAuCN is calculated to be 39.7±0.5 kJ mol⁻¹ at 273 K. To the best of our knowledge this is the first $\Delta G^{\#}$ reported for a scrambling reaction described by (1). The data presented indicate that ligand



Figure 4 Plot of ¹³C vs ¹⁵N chemical shifts of R_3 PAuCN complexes. The solid lines are regression lines. The ¹³C chemical shift values are taken from ref. 4.

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scrambling can be followed using ¹⁵N NMR spectroscopy. The ¹⁵N chemical shift range for RSAuC¹⁵N⁻ and R₃PAuC¹⁵N is 260 to 265 ppm in D₂O or CD₃OD. We have also demonstrated linear relationship between ¹⁵N chemical shifts of R₃PAuC¹⁵N vs v(CO), which shows that when the basicity of phosphines increases the ¹⁵N chemical shift also increases.

Acknowledgements

This work was supported by the KFUPM Research Committee (project no. CY/CYANIDE/175). We would like to thank Prof. C.F. Shaw III, Chemistry Department, University of Wisconsin, U.S.A., for helpful discussion.

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